RULES COMMITTEE

L. Wiesner, Chairman

Three proposals for changing the "Rules for Testing Seeds" were received. These proposals and one other which has been received earlier were reviewed by the Committee. After committee review, several changes were published for consideration by the Association at the 1977 meeting. These changes were mailed by the Secretary at least 90 days in advance of the meeting.

Because of the nature of several of the proposed changes, the "Rules for Testing Seeds" was not updated to include the changes passed in 1976. Consequently, the Committee would plan to update the Rules after the 1977 meeting. This Committee should also review Appendix I "Seedling Descriptions."

The AOSA has received correspondence from several individuals concerning the changing of the blowing point for Merion Kentucky bluegrass. In order to explain the Association's actions leading to this decision, a letter was prepared by the Committee Chairman and sent to President Bill Dale.

Committee Members:
L. Everson (1977)
E. Hardin (1979)
H. Smith (1980)
A. Ednie (1981)
L. Wiesner (1978)

The following "Rules" changes were adopted by the AOSA membership at the 1977 Annual Meeting in Amherst, Massachusetts. The effective date for these changes will be October 1, 1977.

REVISION OF "RULES" SECTIONS 4.8(b), 4.9(g and h), 2.6(b), 2.7(g), 2.12(b), 4.2(a and e), and Table 5

1. Section 4.8(b), p. 31:

   Bahiagrass (Paspalum notatum) - all cultivars except Pensacola: Remove the enclosing structures (glumes, lemma and palea) from the caryopsis with the aid of a sharp scalpel. If the seed is fresh or dormant, scratch the surface of the caryopsis lightly.

2. Section 4.9(g), p. 33: Existing sections g & h would become sections i and j:

   Ethephon - a 0.0029% solution of ethephon {(2-chloroethyl) phosphonic acid} shall be used to moisten the substratum. This solution is prepared by mixing 0.6 ml of a stock solution containing 2 lbs. active material per gallon in a propylene glycol

base with 5,000 ml of distilled water. A solution which is five times the normal concentration (0.0029) may be used for extremely dormant seeds, provided seeds are transferred to substratum moistened with water after 1 to 3 days.

Section 4.9(h), p. 33:

Ethylene - Five (5) ml of ethylene gas per cubic foot of germinator space are injected into a germinator in which peanut seeds in moist rolled towels have been placed. Following injection of ethylene, the germinator is kept closed until the first count (5 days). If germinator door is opened for the purpose of checking or re-wetting the samples, another injection at the same rate may be made.

Table 3, p. 35: (Change is underlined):

	Substrata	Temp.	First Count	Final Count	additional directions Specific Requirements	Fresh and Dormant Seed
Arachis hypogaea Peanut	B,T.S.	20-30,	5	10	Remove shells; Photos 19541, 19542	Ethephon, ethylene (refer to Sec. 4.9g and h

3. Table 5. Tree and Shrub Seeds, pp. 69-76: (Change is underlined):

Kind of seed	Substrata	Temp.	Test Duration days	Additional directions
Picea glauca white spruce	TB	20-30	21	Light; some Canadian seed sources require prechill for 14-21 days at 3-5C
Pinus strobus eastern white pine	TB,P	20-30	21	Light; more than 8 hr. light may be beneficial to some lots, sensitive to drying;prechill 28-42 days at 3-5C
	P	22	28	Light for 16 hr; prechill 28-42 days at 3-5C
Pinus sylvestris scotch pine	TB,P	20-30	14	Light; seed from eastern Mediterranean (Turkey, Greece, Bulgaria) provinces may require prechill 21 days at 3-5C

4. Section 2.6b(3):

Entire spikelets in Agrostis, Panicum, and Setaria. Entire spikelets which may have attached rachis segments, pedicels and sterile spikelets in Andropogon, Sorghum, and Sorghastrum.

5. Section 2.6b(8):

Single units as defined in Section 2.12b.

6. Section 2.7g:

Seed units of the grass family listed in section 2.6b(1) through (5) provided a caryopsis with some degree of endosperm development can be detected in the unit either by slight pressure or by examination over light. Refer to 2.7g(1) and (2) for species in which determination of endosperm development is not necessary. Refer to sections 2.7h and 2.10a(8) when nematode galls, fungus bodies, etc. have replaced the caryopsis in seed units.

7. Section 2.7g(1):

Spikelet, spikelet groups, and multiple forets (double florets) of side-oats grama (Bouteloua curtipendula); multiple florets (double florets) of blue grama (Bouteloua gracilis); spikelet groups and spikelets with attached structures of little bluestem (Andropopgon scoparius); and intact burs of buffalograss (Buchloe dactyloides); shall be considered pure seed whether or not a caryopsis is present. Refer to section 2.10a(9) for the classification of burs which are visibly empty. (See Notice at the end of this report)

8. Section 2.12b:

Definition: A multiple unit is a seed unit that includes at least one fertile floret plus one or more of attached structures as follows:

1. a sterile floret that extends to or beyond the tip of the fertile floret;

2. basally attached glume, glumes, or sterile florets of any length.

The length of an awn shall be disregarded when determining the length of fertile floret or an attached structure. Any seed unit without attached structures, as described above, shall be considered a single unit.

9. Section 4.2a:

Seed germination - In seed laboratory practice, germination is defined as the emergence and development from the seed embryo

of those essential structures which, for the kind of seed in question, are indicative of the ability to produce a normal plant under favorable conditions. Refer to 4.2d and e.

10. Section 4.2e:

Dormant seed - Viable seeds, other than hard seeds, which fail to germinate when provided the specified germination conditions for the kind of seed in question. Viability of ungerminated seeds may be determined by any appropriate method or combination of methods, such as a cutting test, tetrazolium test, scarification, and application of germination promoting chemicals. The percentage dormant seed, if present, shall be reported in addition to the percentage germination for the following species: bahiagrass (Paspalum notatum), bluestems (Andropogon spp.), buffalograss (Buchloe dactyloides), buffelgrass (Pennisetum ciliare), gramas (Bouteloua spp.), green needlegrass (Stipa viridula), indian ricegrass (Oryzopsis hymenoides), lovegrass (Eragrostis spp.), sand dropseed (Sporobolus cryptandrus), smilo (Oryzopsis miliacea), switchgrass (Panicum virgatum), veldtgrass (Ehrharta calycina), western wheatgrass (Agropyron smithii), and yellow indiangrass (Sorghastrum nutans).

(NOTE: An appropriate footnote will be inserted in germination table 3 on the above species).

NOTICE

SUBJECT: Converting to the new method for testing blue grama, side-oats, grama, and little bluestem.

A new method for testing blue grama, side-oats grama and little bluestem has been officially accepted at the Amherst meetings of AOSA. These new procedures will be in effect on October 1, 1977. Conversion to the new method should begin now for seed to be sold this coming year.

When analysis for both purity and germination are requested, little or no problems are anticipated. When analysis for germination only are requested, a determination of purity (at least for pure seed content) must be made. The new method raises the pure seed content substantially over the pure seed content obtained by the old method and correspondingly lowers the germination. When converting to the new method, both corrections are necessary.

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